

This is the mountant we hand out free to the Facility users either with or without DAPI as counter stain. It hardens so it creates a semi permanent mounted slide. However, please bear in mind:

- 1) that Mowiol (also referred as PVA) refractive index changes as it polymerizes. Therefore it is not a good mounting medium for deconvolution work.
- 2) that Mowiol shrinks substantially as it polymerizes and it is not suitable for thick specimen (>30um or so).

Important: As Mowiol shrinks considerably, it is critical apply enough Mowiol to avoid formation of air bubbles. For single coverglass 22x22mm with adherent cells, first, apply one drop of Mowiol onto a slide. Then, blot off buffer solution from the coverglass as possible without let it dries. Then put the coverglass with cell side facing the slide onto the drop of Mowiol. Apply very gentle pressure on the coverglass so Mowiol spread but not enough force to squeeze out the Mowiol. Leave the slide at room temperature covered in dark for 2hrs to polymerize. Increase the amount of Mowiol for large coverglasses.

Mowiol Recipe:

Mowiol is in powder form, also known as poly vinyl alcohol. It is available from

Calbiochem (Mowiol 4-88 catalogue number 475904).

Recipe –found in Harlow and Lane’s “Using Antibodies”

Add 2.4g Mowiol to 6g glycerol and stir to mix. Add 6ml of water and stir for several hours at room temperature. Add 12 ml of 0.2M phosphate buffer, ph 7.4. and heat to 50°C for 30min with constant mixing. After the Mowiol dissolves, (probably won’t completely dissolve), allow the mixture to cool, and add 0.1% n-propyl gallate (Sigma P3130), then clear by centrifugation at 5000xg for 15 min. Aliquot in cryovials and store at –20°C. Stock solutions are stable for up to 2 weeks at +4°C.